#### [Cat. No.] K-7170

#### Introduction

AccuRapid™ TA Cloning Kit is designed for directional cloning of PCR products amplified with Taq DNA polymerase into linearized T-vectors without any restriction sites or modifications. Tag DNA polymerase adds a single nucleotide, deoxyadenosine (dA) to the 3'-end of the amplified PCR products. The linearized T-vector has a single deoxythymidine (dT) at the 3'-end. The TA Cloning allows the hybridization of the complementary 3'-end A overhangs of PCR products and 3'-end T overhangs of linearized T-vector.

#### Features & Benefits

- Convenient: Easy excision of DNA insert using BamH I and M13 primer sites for sequencing.
- Rapid: 15 min reaction, enables cloning of various sizes of insert DNA.
- Blue/white screening: Provides blue/white selection function through LacZα gene.

#### Components

Storage Store at -20°C.

Components	Amount
pBHA-T vector (25 ng/µl)	40 µl
Control Insert (70 ng/µl)	20 μΙ
AccuRapid™ 2X Reaction buffer	100 μΙ
T4 DNA Ligase (200 U/μI)	20 μΙ

<sup>\*</sup> Note: For research use only. Not for use in diagnostic or therapeutic procedures.

#### Online Resources





Korean

Visit our product page for additional information and protocols.

## Ordering Information

Description	Cat. No.
AccuRapid™ TA Cloning Kit	K-7170

### Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products. services, specifications or product descriptions at any time without notice

# **Explanation of Symbols**









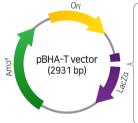








# pBHA-T Vector Map



M13 reverse primer →lacZa ATG CAC AGG AAA CAG CTA TGA CCA TGA TTA CGC CAA GCT ATT TAG GTG ACA CTA TAG AAT ACT GTG TCC TTT GTC GAT ACT GGT ACT AAT GCG GTT CGA TAA ATC CAC TGT GAT ATC TTA TGA BamHI CAA GCT ATG CAT CAA GCT TGG TAC CGA GCT CGG ATC CAC TAG TAA CGG CCG CCA GTG TGC GTT CGA TAC GTA GTT CGA ACC ATG GCT CGA GCC TAG GTG ATC ATT GCC GGC GGT CAC ACG BamH I TGG AAT TCT GCA GAT CCA ATA CT PCR Product AGT ATT GGG GGA TCC ATC CAT CAC ACT ACC TTA AGA CGT CTA GGT TAT GT TGA Ngt | Xho | Xba | T7 promoter GGC GGC CGC TCG AGC ATG CAT CTA GAG GGC CCA ATT CGC [CCT ATA GTG AGT CGT ATT ALX CCC CCG GG AGC TCG TAC GTA GAT CTC CCG GGT TAA GCC [GGA TAT CAC TCA GCA TAA TST M13 forward primer ATT CAC TGG CCG TCG TTT TAC AAC GTC GTG ACT GGG AAA ACC TAA GTG ACC GGC AGC AAA ATG TTG CAG CAC TGA CCC TTT TGG

pBHA-T Vector: 2,931 bp

M13 Reverse primer site: bases 1-17 LacZα gene: bases 13-441

SP6 promoter: bases 35-53 T7 promoter: bases 236-254 M13 Forward primer site: bases 261-277 Ampicillin resistance gene: bases 1,003-1,863 pUC ori (high copy number): bases 2,034-2,622

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# **Experimental Procedures**

	Steps	Procedure Details				
1 TA	Preparation of PCR products	1. Perform PCR with <i>Taq</i> DNA polymerase.  * Note: For efficient A-tailing of PCR products, it is recommended to proceed with final extension at 72°C for more than 5 min.  2. Confirm the amplified PCR products by agarose gel electrophoresis.  3. Purify the PCR products using a gel purification kit.  4. Quantify the purified PCR products to use ligation reaction.				
2	Thawing materials	Take out the pBHA-T vector and <i>AccuRapid</i> ™ 2X Reaction buffer from the kit and thaw them on ice.				
3		6. Prepare the ligation reaction mixture.     Preparation of ligation reaction mixture     Components Positive Sample				
	Preparation of ligation reaction mixture	pBHA-T vector (25 ng/µl) PCR products Control Insert (70 ng/µl) AccuRapid™ 2X Reaction buffer T4 DNA Ligase (200 U/µl) Nuclease-free water Total volume * Note: In general, 0.5-2 µl of PCR product is used vector) or higher for a more efficient ligation react 7. Mix the reaction mixture by tapping and	2 µl  - 1 µl 5 µl 1 µl 1 µl 10 µl 1, and it is recommended to add	2 µl 0.5-2 µl* - 5 µl 1 µl Variable 10 µl		
	Perform transformation using heat-shock or electroporation					
4	Ligation and transformation	Incubate the ligation reaction mixture for at least 15 min at 25°C. For large amounts of transformants, incubate the reaction mixture for 1 hr.      Spread the transformation mixture on the LB agar plates containing ampicillin, X-Gal, and IPTG.				
5	Analysis	Pick at least 10 white colonies and perform colony PCR.     Analyze the bands using electrophoresis.				