

[Cat. No.] **K-2051**

Introduction

AccuPower® CycleScript™ RT Master Mix is applied with BIONEER's patent technology called Cyclic Temperature Reverse Transcription (CTRT), which not only increases the efficiency, but also is effective for full-length cDNA synthesis. The CTRT reaction is composed of 2 or 3 steps. The step 1 is performed at 15-25°C, at which short primers are fully annealed. Then, the step 2 is performed at 42-48°C for cDNA synthesis. The step 3 is performed at high temperature 50-55°C at which secondary structure of template RNA obstructing reverse transcription is released. This product is a ready-to-use mixture containing thermostable CycleScript™ Reverse Transcriptase, reaction buffer, DTT, dNTPs, and stabilizer. Primers (Oligo dT₂₀ and Oligo dN₆) are provided in separated tubes. It simplifies preparation of reverse transcription reaction mixture by adding template RNA, primers, and nuclease-free water without any extra process.

Applications

- Sequencing single and double-strand DNA or RNA
- Random priming reaction
- cDNA library construction
- Probe labeling
- mRNA 5'-end mapping by primer extension analysis
- PCR
- Real-time PCR

Features & Benefits

- Flexible conditions: Cyclic Temperature Reverse Transcription (CTRT) is more efficient cDNA synthesis by using higher temperature than the conventional methods using at 42°C. Primer annealing is performed at a range of low temperature (15-40°C) and secondary structures of template is released by repeating the step 2 or 3 at a range of high temperature (50-55°C).
- Stability: Included stabilizer and thermostable reverse transcriptase allow to react at a higher temperature up to 55°C.
- Controllable reaction time: Reaction time can be controlled depending on the number and size of copies of target gene. In case of high-copy gene, cDNA can be synthesized even with 10 minutes reverse transcription reaction.
- Reproducibility: Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.
- Ease-of-use: Reactants are included in a tube, it allows any user simply perform cDNA synthesis by adding template RNA and nuclease-free water.

Components

Components	Amount
2X Master Mix	1 ml
Oligo dT ₂₀ (100 pmol/μl)	100 μl
Oligo dN ₆ (100 pmol/μl)	100 μl

Composition

Composition	Concentration
CycleScript™ Reverse Transcriptase	200 U
5X Reaction buffer	1X
DTT	0.25 mM
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 μM
RNase inhibitors	1 U

Specifications

CycleScript™ Reverse Transcriptase	
DNase activity	No
RNase activity	No
Fragment size	Up to 9 kb

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

Online Resources



Korean



English

Visit our **product page** for additional information and protocols

Ordering Information

Description	Cat. No.
1 ml of 2X Master Mix solution	1 ml x 1 ea
	K-2051

* **Note:** For Master Mix products, primers (dT₂₀ & dN₆) are provided in a separate tube.

Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

Explanation of Symbols



Batch Code



Biological Risks



Catalog Number



Caution



Consult Instructions For Use



Contains Sufficient for <n> tests



Do not Re-use



Manufacturer



Research Use Only



Temperature Limitation



Use-by Date

Experimental Procedures

Steps		Procedure Details																																																
1	 Thaw reagents	1. Thaw <i>AccuPower® CycleScript™</i> RT Master Mix on ice and mix thoroughly before use. Then, briefly spin down components. 2. Dispense appropriate volumes of <i>AccuPower® CycleScript™</i> RT Master Mix into PCR tubes (not provided). Use 10 µl and 25 µl of 2X Master Mix for 20 µl reaction and 50 µl reaction, respectively.																																																
2	 Preparation of reaction mixture	3. Add template RNA, primers, and nuclease-free water into PCR tubes containing <i>AccuPower® CycleScript™</i> RT Master Mix. <ul style="list-style-type: none"> Amount of template RNA and primers <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th colspan="2">Components</th> <th>20 µl reaction</th> <th>50 µl reaction</th> </tr> </thead> <tbody> <tr> <td rowspan="2">Template RNA</td> <td>Total RNA</td> <td>0.1-1.0 µg</td> <td>0.1-1.0 µg</td> </tr> <tr> <td>Poly(A) RNA</td> <td>0.01-1.0 µg</td> <td>0.01-1.0 µg</td> </tr> <tr> <td rowspan="3">Primers</td> <td>Oligo dT</td> <td>50-100 pmol</td> <td>250 pmol</td> </tr> <tr> <td>Random primer</td> <td>100 pmol</td> <td>250 pmol</td> </tr> <tr> <td>Gene specific primer</td> <td>10-50 pmol</td> <td>10-50 pmol</td> </tr> </tbody> </table> 4. Mix the reaction mixture by pipetting or vortexing, and briefly spin down.	Components		20 µl reaction	50 µl reaction	Template RNA	Total RNA	0.1-1.0 µg	0.1-1.0 µg	Poly(A) RNA	0.01-1.0 µg	0.01-1.0 µg	Primers	Oligo dT	50-100 pmol	250 pmol	Random primer	100 pmol	250 pmol	Gene specific primer	10-50 pmol	10-50 pmol																											
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	 Option	<ul style="list-style-type: none"> If PCR is needed, transfer 2-5 µl of reaction mixture (synthesized cDNA) into <i>AccuPower®</i> PCR PreMix tubes (Cat. No. K-2012, not provided), and perform the reaction under the following conditions. <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td>95°C</td> <td>5 min</td> <td>1 cycle</td> </tr> <tr> <td>Denaturation</td> <td>95°C</td> <td>20 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td>45-65°C</td> <td>20 sec</td> <td>25-35 cycles</td> </tr> <tr> <td>Extension</td> <td>72°C</td> <td>0.5-1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td>72°C</td> <td>3-5 min</td> <td>1 cycle</td> </tr> </tbody> </table> <p>* Note: For maximum yield and specificity, temperatures and cycling times should be optimized for each new template DNA or primers.</p>	Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	20 sec		Annealing	45-65°C	20 sec	25-35 cycles	Extension	72°C	0.5-1 min/kb		Final extension	72°C	3-5 min	1 cycle																								
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