

[Cat. No.] Please refer to the **Ordering Information**

Introduction

AccuPower® HotStart PCR PreMix is based on a concept chemical interaction between pyrophosphate (PPi) and pyrophosphatase (PPase). DNA polymerase is required Mg²⁺ to activate, but included PPi binds with high affinities to Mg²⁺ resulting in inhibition of polymerase activity. When the temperature rises during denaturation step, Mg-PPi complex is decomposed into 2Pi and Mg²⁺ by PPase. Then, activated DNA polymerase proceed reactions. This prevents the formation of mis-primed products and primer-dimers during the reaction set up process resulting in improved PCR specificity. This product contains vacuum-dried components including HotStart DNA Polymerase, dNTPs, reaction buffer, stabilizer, and tracking dye. It simplifies preparation of reaction mixture by adding template DNA and primers without any extra process. After the reaction, samples can be applied directly on agarose gel for analysis.

Applications

- High specificity PCR
- High sensitivity PCR
- gDNA template PCR
- Low-copy target PCR
- Multiple primer pair PCR
- cDNA template PCR

Features & Benefits

- **Specificity:** Minimized non-specific amplification and maximized PCR efficiency by using BIONEER's differentiated PyroHotstart technology.
- **Sensitivity:** Excellent sensitivity and amplification efficiency even with small amounts of DNA.
- **Stability:** Included stabilizer enables delivery at room temperature and provides increased stability compared to solution-type products.
- **User-friendly:** Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform PCR by adding template DNA and primers.
- **Reproducibility:** Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.

Composition

Composition	Concentration
HotStart DNA Polymerase	1 U
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 µM
Reaction buffer with 1.5 mM MgCl ₂	1X
Stabilizer and tracking dye	O

Specifications

HotStart DNA Polymerase	
5' to 3' exonuclease activity	No
3' to 5' exonuclease activity	No
3'-A overhang	Yes
Fragment size	Up to 12 kb

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

Precautions

- This enzyme is specifically optimized for increasing base incorporation rate by inactivating 5' to 3' exonuclease activity. Therefore, this product is not recommended to use for real-time PCR using hydrolysis probe method.

Online Resources



Korean



English

Visit our **product page** for additional information and protocols

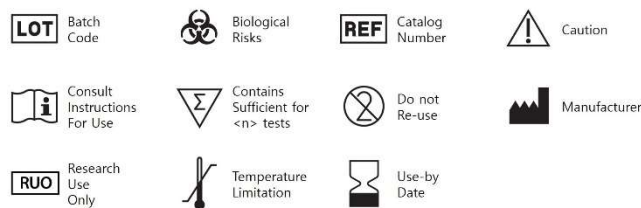
Ordering Information

Description	Cat. No.
0.2 ml thin-wall 96 tubes	20 µl/rxn K-5050
8-tube strips with attached cap	50 µl/rxn K-5052
480 tubes	20 µl/rxn K-5051
	50 µl/rxn K-5057
0.5 ml thin-wall tubes with attached cap	100 tubes 50 µl/rxn K-5053



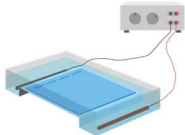
Notice

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Explanation of Symbols



Experimental Procedures

Steps		Procedure Details																								
1	 Preparation of reaction mixture	<p>1. Add template DNA, primers, and nuclease-free water into <i>AccuPower®</i> Hotstart PCR PreMix tubes to make a total volume of 20 µl or 50 µl. Do not include the dried pellet.</p> <ul style="list-style-type: none"> Preparation of reaction mixture <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Components</th> <th style="text-align: center;">20 µl reaction</th> <th style="text-align: center;">50 µl reaction</th> </tr> </thead> <tbody> <tr> <td>Template DNA (≤ 100 ng)</td> <td style="text-align: center;">Variable (1-10 µl)</td> <td style="text-align: center;">Variable (1-25 µl)</td> </tr> <tr> <td>Forward primer (5 pmol/µl)</td> <td style="text-align: center;">1-2 µl</td> <td style="text-align: center;">2-5 µl</td> </tr> <tr> <td>Reverse primer (5 pmol/µl)</td> <td style="text-align: center;">1-2 µl</td> <td style="text-align: center;">2-5 µl</td> </tr> <tr> <td>Nuclease-free water</td> <td style="text-align: center;">Variable</td> <td style="text-align: center;">Variable</td> </tr> <tr> <td>Total volume</td> <td style="text-align: center;">20 µl</td> <td style="text-align: center;">50 µl</td> </tr> </tbody> </table> <p>2. Dissolve the vacuum-dried blue pellet by vortexing or pipetting, and briefly spin down.</p>	Components	20 µl reaction	50 µl reaction	Template DNA (≤ 100 ng)	Variable (1-10 µl)	Variable (1-25 µl)	Forward primer (5 pmol/µl)	1-2 µl	2-5 µl	Reverse primer (5 pmol/µl)	1-2 µl	2-5 µl	Nuclease-free water	Variable	Variable	Total volume	20 µl	50 µl						
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2	 Incubate reactions in a thermal cycler	<p>3. Perform the reaction under the following conditions.</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Step</th> <th style="text-align: center;">Temperature</th> <th style="text-align: center;">Time</th> <th style="text-align: center;">Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">94°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">94°C</td> <td style="text-align: center;">30 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td style="text-align: center;">50-68°C†</td> <td style="text-align: center;">0.5-1 min</td> <td style="text-align: center;">25-35 cycles</td> </tr> <tr> <td>Extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>* Note: For maximum yield and specificity, temperatures and cycling times should be optimized for each new template DNA or primers. † Set the annealing temperature to 3-5 degrees lower than the T_m of the primers.</p>	Step	Temperature	Time	Cycles	Pre-denaturation	94°C	5 min	1 cycle	Denaturation	94°C	30 sec		Annealing	50-68°C†	0.5-1 min	25-35 cycles	Extension	72°C	1 min/kb		Final extension	72°C	5 min	1 cycle
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3	 Analyze with gel electrophoresis	<p>4. After the reaction, maintain the reaction mixture at 4-8°C. The samples can be stored at -20°C until use.</p> <p>5. Load 5 µl of samples on agarose gel without adding a loading-dye mixture, and perform gel electrophoresis for analysis.</p>																								