

[Cat. No.] Please refer to the **Ordering Information**

Introduction

AccuPower® ProFi Taq PCR PreMix is ideal for convenient and high fidelity of DNA amplification. Included ProFi Taq DNA Polymerase is a unique recombinant Taq DNA polymerase that provides accurate amplification of long template DNA with high fidelity and high efficiency. This product contains vacuum-dried components including ProFi Taq DNA Polymerase, dNTPs, reaction buffer, stabilizer, and tracking dye. It simplifies preparation of reaction mixture by adding template DNA and primers without any extra process. After the reaction, samples can be applied directly on agarose gel for analysis.

Applications

- Primer extension
- Long-range amplification from genomic DNA
- High amplification efficiency
- Excellent performance on difficult templates
- Amplification of low-copy targets
- High yield and high sensitivity PCR

Features & Benefits

- High Efficiency and high sensitivity: Guaranteed accurate PCR products with excellent sensitivity and amplification efficiency even with small amounts of DNA.
- Long-range PCR: Effective amplification of large genomic DNA fragments up to 20 kb of human DNA.
- User-friendly: Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform PCR by adding template DNA and primers.
- Stability: Included stabilizer enables delivery at room temperature and provides increased stability compared to solution-type products.
- Reproducibility: Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.

Composition

Composition	Concentration
ProFi Taq DNA Polymerase	1 U
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 µM
Reaction buffer with 1.5 mM MgCl ₂	1X
Stabilizer and tracking dye	0

Specifications

ProFi Taq DNA Polymerase	
5' to 3' exonuclease activity	Yes
3' to 5' exonuclease activity	Yes
3'-A overhang	Yes
Fragment size	Up to 20 kb

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

Online Resources



Korean



English

Visit our **product page** for additional information and protocols

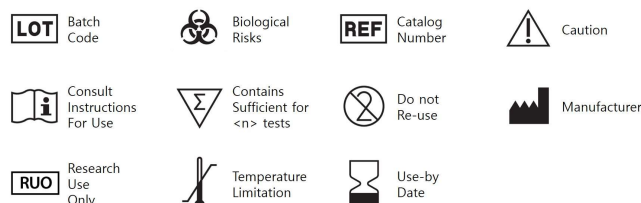
Ordering Information

Description	Cat. No.
0.2 ml thin-wall 96 tubes	K-2631
8-tube strips with attached cap	K-2632
50 µl/rxn	K-2633
20 µl/rxn	K-2634



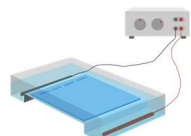

Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

Explanation of Symbols



Experimental Procedures

Steps		Procedure Details																								
1	 Preparation of reaction mixture	<p>1. Add template DNA, primers, and nuclease-free water into <i>AccuPower® ProFi Taq</i> PCR PreMix tubes to make a total volume of 20 µl or 50 µl. Do not include the dried pellet.</p> <ul style="list-style-type: none"> Preparation of reaction mixture <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Components</th> <th style="text-align: center;">20 µl reaction</th> <th style="text-align: center;">50 µl reaction</th> </tr> </thead> <tbody> <tr> <td>Template DNA (1-500 ng)</td> <td style="text-align: center;">Variable (1-10 µl)</td> <td style="text-align: center;">Variable (1-25 µl)</td> </tr> <tr> <td>Forward primer (10 pmol/µl)</td> <td style="text-align: center;">0.5-2 µl</td> <td style="text-align: center;">1-5 µl</td> </tr> <tr> <td>Reverse primer (10 pmol/µl)</td> <td style="text-align: center;">0.5-2 µl</td> <td style="text-align: center;">1-5 µl</td> </tr> <tr> <td>Nuclease-free water</td> <td style="text-align: center;">Variable</td> <td style="text-align: center;">Variable</td> </tr> <tr> <td>Total volume</td> <td style="text-align: center;">20 µl</td> <td style="text-align: center;">50 µl</td> </tr> </tbody> </table> <p>2. Dissolve the vacuum-dried blue pellet by vortexing or pipetting, and briefly spin down.</p>	Components	20 µl reaction	50 µl reaction	Template DNA (1-500 ng)	Variable (1-10 µl)	Variable (1-25 µl)	Forward primer (10 pmol/µl)	0.5-2 µl	1-5 µl	Reverse primer (10 pmol/µl)	0.5-2 µl	1-5 µl	Nuclease-free water	Variable	Variable	Total volume	20 µl	50 µl						
Components	20 µl reaction	50 µl reaction																								
Template DNA (1-500 ng)	Variable (1-10 µl)	Variable (1-25 µl)																								
Forward primer (10 pmol/µl)	0.5-2 µl	1-5 µl																								
Reverse primer (10 pmol/µl)	0.5-2 µl	1-5 µl																								
Nuclease-free water	Variable	Variable																								
Total volume	20 µl	50 µl																								
2	 Incubate reactions in a thermal cycler	<p>3. Perform the reaction under the following conditions.</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Step</th> <th style="text-align: center;">Temperature</th> <th style="text-align: center;">Time</th> <th style="text-align: center;">Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">15-20 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td style="text-align: center;">45-65°C*</td> <td style="text-align: center;">15-30 sec</td> <td style="text-align: center;">25-35 cycles</td> </tr> <tr> <td>Extension</td> <td style="text-align: center;">68°C</td> <td style="text-align: center;">1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td style="text-align: center;">68°C</td> <td style="text-align: center;">3-5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>* Optimal annealing temperature depends on the melting temperature of the primers.</p>	Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	15-20 sec		Annealing	45-65°C*	15-30 sec	25-35 cycles	Extension	68°C	1 min/kb		Final extension	68°C	3-5 min	1 cycle
Step	Temperature	Time	Cycles																							
Pre-denaturation	95°C	5 min	1 cycle																							
Denaturation	95°C	15-20 sec																								
Annealing	45-65°C*	15-30 sec	25-35 cycles																							
Extension	68°C	1 min/kb																								
Final extension	68°C	3-5 min	1 cycle																							
3	 Analyze with gel electrophoresis	<p>4. After the reaction, maintain the reaction mixture at 4-8°C. The samples can be stored at -20°C until use.</p> <p>5. Load samples on agarose gel without adding a loading-dye mixture, and perform gel electrophoresis for analysis.</p>																								
	 Option	<ul style="list-style-type: none"> If primer's T_m value is more than 65°C or PCR product size is more than 10 kb, follow the conditions as below. <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="text-align: center;">Step</th> <th style="text-align: center;">Temperature</th> <th style="text-align: center;">Time</th> <th style="text-align: center;">Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">5 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">15-20 sec</td> <td></td> </tr> <tr> <td>Annealing/Extension</td> <td style="text-align: center;">68°C</td> <td style="text-align: center;">1 min/kb*</td> <td style="text-align: center;">30-35 cycles</td> </tr> <tr> <td>Final Extension</td> <td style="text-align: center;">68°C</td> <td style="text-align: center;">3-5 min</td> <td style="text-align: center;">1 cycle</td> </tr> </tbody> </table> <p>* Annealing/Extension time depends on target length. Perform 15 min for 20 kb and 20 min for 30 kb.</p>	Step	Temperature	Time	Cycles	Pre-Denaturation	95°C	5 min	1 cycle	Denaturation	95°C	15-20 sec		Annealing/Extension	68°C	1 min/kb*	30-35 cycles	Final Extension	68°C	3-5 min	1 cycle				
Step	Temperature	Time	Cycles																							
Pre-Denaturation	95°C	5 min	1 cycle																							
Denaturation	95°C	15-20 sec																								
Annealing/Extension	68°C	1 min/kb*	30-35 cycles																							
Final Extension	68°C	3-5 min	1 cycle																							