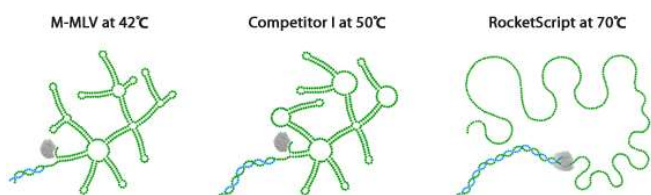


[Cat. No.] **K-2105**

### Introduction

AccuPower<sup>®</sup> RocketScript<sup>™</sup> RT Master Mix can effectively synthesize cDNA with complex secondary RNA structures by using RocketScript<sup>™</sup> Reverse Transcriptase developed by BIONEER. RocketScript<sup>™</sup> Reverse Transcriptase originated and engineered from M-MLV Reverse Transcriptase can provide increased thermal stability resulting in effective full length first strand cDNA synthesis. It can be used at various temperatures (42-70°C) allowing increased specificity, higher yields of cDNA, and more full-length product than other reverse transcriptase. This product is a ready-to-use mixture for cDNA synthesis including RocketScript<sup>™</sup> Reverse Transcriptase, reaction buffer, DTT, dNTPs, and RNase inhibitor. It simplifies preparation of reverse transcription reaction mixture by adding template RNA and primers without any extra process.



**Figure 1. Schematic diagram of 5' UTR of a gene, with complex secondary structure, at three different temperatures.**

RocketScript<sup>™</sup> Reverse Transcriptase is fully active at 70°C, so it can synthesize complete gene sequences that M-MLV and other reverse transcriptase can not synthesize.

### Applications

- First-strand synthesis of cDNA from RNA molecules (RT)
- Random priming reaction
- cDNA library construction
- Probe labeling
- mRNA 5'-end mapping by primer extension analysis
- PCR
- Real-time PCR

### Features & Benefits

- **Thermostable activity:** RocketScript<sup>™</sup> Reverse Transcriptase has excellent thermal resistance even at 70°C, so it is suitable for cDNA synthesis of secondary RNA structures. The cDNA synthesis temperatures can be set at various temperatures (42-70°C) according to user's experimental conditions, allowing effective cDNA synthesis.
- **Stability:** Included stabilizer and RNase inhibitor provides high resistance to degradation.
- **Reproducibility:** Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.
- **Ease-of-use:** Reactants are included in a tube, it allows any user simply perform cDNA synthesis by adding template RNA and primers.

### Composition

Composition	Concentration
RocketScript <sup>™</sup> Reverse Transcriptase	200 U
5X Reaction buffer	1X
DTT	0.25 mM
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 μM
RNase inhibitor	1 U

### Specifications

RocketScript <sup>™</sup> Reverse Transcriptase	
DNase activity	No
RNase activity	No
Fragment size	Up to 10 kb

### Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

### Online Resources



Korean



English

Visit our **product page** for additional information and protocols

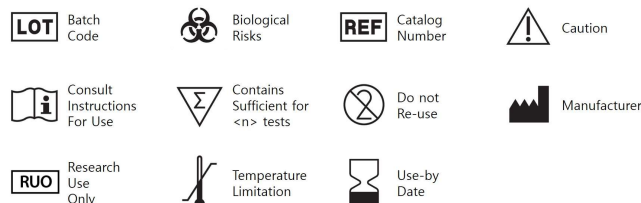
### Ordering Information

Description	Cat. No.
1 ml of 2X Master Mix solution	1 ml x 1 ea K-2105





### Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

### Explanation of Symbols



**Experimental Procedures**

Steps		Procedure Details																																					
1	 <b>Thaw reagents</b>	<p>1. Thaw <i>AccuPower® RocketScript™</i> RT Master Mix on ice and mix thoroughly before use. Then, briefly spin down components.</p> <p>2. Dispense appropriate volumes of <i>AccuPower® RocketScript™</i> RT Master Mix into PCR tubes (not provided). Use 10 µl and 25 µl of 2X Master Mix for 20 µl reaction and 50 µl reaction, respectively.</p>																																					
2	 <b>Preparation of reaction mixture</b>	<p>3. Add template RNA and primers into PCR tubes containing <i>AccuPower® RocketScript™</i> RT Master Mix, and then fill up the nuclease-free water to make a total volume of 20 µl or 50 µl.</p> <ul style="list-style-type: none"> <li>Amount of template RNA and primers</li> </ul> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th colspan="2">Components</th> <th>20 µl reaction</th> <th>50 µl reaction</th> </tr> </thead> <tbody> <tr> <td rowspan="2" style="text-align: center;">Template RNA</td> <td>Total RNA</td> <td>0.01-5 µg</td> <td>0.01-5 µg</td> </tr> <tr> <td>Poly(A) RNA</td> <td>0.01-5 µg</td> <td>0.01-5 µg</td> </tr> <tr> <td rowspan="3" style="text-align: center;">Primers</td> <td>  Oligo dT</td> <td>100 pmol</td> <td>100 pmol</td> </tr> <tr> <td>  Random primer</td> <td>100 pmol</td> <td>100 pmol</td> </tr> <tr> <td>  Gene specific primer</td> <td>10-50 pmol</td> <td>10-50 pmol</td> </tr> </tbody> </table> <p>4. Mix the reaction mixture by pipetting or vortexing, and briefly spin down.</p>	Components		20 µl reaction	50 µl reaction	Template RNA	Total RNA	0.01-5 µg	0.01-5 µg	Poly(A) RNA	0.01-5 µg	0.01-5 µg	Primers	Oligo dT	100 pmol	100 pmol	Random primer	100 pmol	100 pmol	Gene specific primer	10-50 pmol	10-50 pmol																
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3	 <b>cDNA synthesis</b>	<p>5. Perform the reaction under the following conditions.</p> <p>5-1. Example 1</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th rowspan="2">Step</th> <th colspan="4">Temperature</th> <th rowspan="2">Time</th> </tr> <tr> <th>dN<sub>6</sub></th> <th>dN<sub>12</sub></th> <th>dT<sub>20</sub></th> <th>Gene specific</th> </tr> </thead> <tbody> <tr> <td>Primer annealing</td> <td>15°C</td> <td>30°C</td> <td>37°C</td> <td>T<sub>m</sub> of primers</td> <td>10 min</td> </tr> <tr> <td>cDNA synthesis</td> <td colspan="3">42°C</td> <td></td> <td>30 min</td> </tr> <tr> <td>Heat inactivation</td> <td colspan="3">95°C</td> <td></td> <td>5 min</td> </tr> </tbody> </table> <p>5-2. Example 2</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> </tr> </thead> <tbody> <tr> <td>cDNA synthesis</td> <td>42-70°C</td> <td>1 hr</td> </tr> <tr> <td>Heat inactivation</td> <td>95°C</td> <td>5 min</td> </tr> </tbody> </table> <p>* <b>Note:</b> For difficult or high GC-content templates, perform cDNA synthesis at 55°C.</p> <p>6. After the reaction, maintain the reaction mixture at 4°C. The samples can be stored at -20°C until use.</p>	Step	Temperature				Time	dN <sub>6</sub>	dN <sub>12</sub>	dT <sub>20</sub>	Gene specific	Primer annealing	15°C	30°C	37°C	T <sub>m</sub> of primers	10 min	cDNA synthesis	42°C				30 min	Heat inactivation	95°C				5 min	Step	Temperature	Time	cDNA synthesis	42-70°C	1 hr	Heat inactivation	95°C	5 min
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 <b>Option</b>	<ul style="list-style-type: none"> <li>If PCR is needed, transfer 2-5 µl of reaction mixture (synthesized cDNA) into <i>AccuPower®</i> PCR PreMix tubes (Cat. No. K-2012, not provided), and perform the reaction under the following conditions.</li> </ul> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td>95°C</td> <td>5 min</td> <td>1 cycle</td> </tr> <tr> <td>Denaturation</td> <td>95°C</td> <td>20 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td>45-65°C</td> <td>20 sec</td> <td>25-35 cycles</td> </tr> <tr> <td>Extension</td> <td>72°C</td> <td>0.5-1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td>72°C</td> <td>3-5 min</td> <td>1 cycle</td> </tr> </tbody> </table> <p>* <b>Note:</b> For maximum yield and specificity, temperatures and cycling times should be optimized for each new template DNA or primers.</p>	Step	Temperature	Time	Cycles	Pre-denaturation	95°C	5 min	1 cycle	Denaturation	95°C	20 sec		Annealing	45-65°C	20 sec	25-35 cycles	Extension	72°C	0.5-1 min/kb		Final extension	72°C	3-5 min	1 cycle														
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