

[Cat. No.] **K-2605, K-2606**

Introduction

AccuPower® Taq PCR PreMix (Negative Dye) is the powerful technology for convenient and easy performance of DNA amplification. This product contains vacuum-dried components including Taq DNA Polymerase, dNTPs, reaction buffer, and stabilizer. It simplifies preparation of reaction mixture by adding template DNA and primers without any extra process. After the reaction, loading-dye mixture must be added to samples, and then loaded on agarose gel for analysis.

Applications

- Conventional PCR
- Primer extension
- TA cloning
- Gene sequencing

Features & Benefits

- **Stability:** Included stabilizer enables delivery at room temperature and provides increased stability compared to solution-type products.
- **Sensitivity:** Excellent sensitivity and amplification efficiency even with small amounts of DNA.
- **User-friendly:** Reactants are individually packaged in each of the PCR tubes, it allows any user simply perform PCR by adding template DNA and primers.
- **Reproducibility:** Mass production under ISO 9001 quality system allows minimized deviation between lots and reproducible results in replicated tests performed under same conditions and variation.

Composition

Composition	Concentration
Taq DNA Polymerase	1 U
dNTPs (dATP, dCTP, dGTP, dTTP)	Each 250 µM
Reaction buffer with 1.5 mM MgCl ₂	1X
Stabilizer	O

Specifications

Taq DNA Polymerase	
5' to 3' exonuclease activity	Yes
3' to 5' exonuclease activity	No
3'-A overhang	Yes
Fragment size	Up to 10 kb

Storage

Store at -20°C. If stored in the recommended temperature, this product will be stable until the expiration date printed out on the label.

Online Resources



Korean



English

Visit our **product page** for additional information and protocols

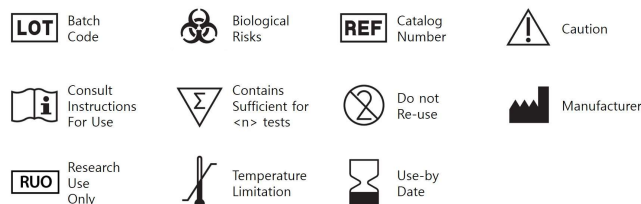
Ordering Information

Description	Cat. No.
0.2 ml thin-wall 8-tube strips with attached cap	K-2605
96 tubes 20 µl/rxn (-dye)	K-2606
480 tubes 20 µl/rxn (-dye)	



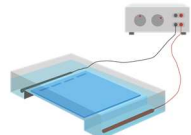

Notice

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Explanation of Symbols



Experimental Procedures

Steps		Procedure Details																									
1	 Preparation of reaction mixture	1. Add template DNA, primers, and nuclease-free water into <i>AccuPower® Taq PCR PreMix (Negative Dye)</i> tubes to make a total volume of 20 µl. Do not include the dried pellet.																									
		<ul style="list-style-type: none"> Amount of template <table border="1"> <thead> <tr> <th>Template DNA</th> <th>Amount of template</th> </tr> </thead> <tbody> <tr> <td>Bacteriophage λ, Plasmid DNA</td> <td>100 fg-200 ng</td> </tr> <tr> <td>Total genomic DNA</td> <td>1-500 ng</td> </tr> </tbody> </table> <ul style="list-style-type: none"> Preparation of reaction mixture <table border="1"> <thead> <tr> <th>Components</th> <th>20 µl reaction</th> </tr> </thead> <tbody> <tr> <td>Template DNA</td> <td>Variable (1-10 µl)</td> </tr> <tr> <td>Forward primer (10 pmol/µl)</td> <td>0.5-2 µl</td> </tr> <tr> <td>Reverse primer (10 pmol/µl)</td> <td>0.5-2 µl</td> </tr> <tr> <td>Nuclease-free water</td> <td>Variable</td> </tr> <tr> <td>Total volume</td> <td>20 µl</td> </tr> </tbody> </table>				Template DNA	Amount of template	Bacteriophage λ, Plasmid DNA	100 fg-200 ng	Total genomic DNA	1-500 ng	Components	20 µl reaction	Template DNA	Variable (1-10 µl)	Forward primer (10 pmol/µl)	0.5-2 µl	Reverse primer (10 pmol/µl)	0.5-2 µl	Nuclease-free water	Variable	Total volume	20 µl				
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2	 Incubate reactions in a thermal cycler	2. Dissolve the vacuum-dried pellet by vortexing or pipetting, and briefly spin down.																									
		3. Perform the reaction under the following conditions. <table border="1"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td>95°C</td> <td>1-5 min*</td> <td>1 cycle</td> </tr> <tr> <td>Denaturation</td> <td>95°C</td> <td>30 sec</td> <td></td> </tr> <tr> <td>Annealing</td> <td>45-65°C†</td> <td>30 sec</td> <td>25-35 cycles</td> </tr> <tr> <td>Extension</td> <td>72°C</td> <td>0.5-1 min/kb</td> <td></td> </tr> <tr> <td>Final extension</td> <td>72°C</td> <td>3-5 min</td> <td>1 cycle</td> </tr> </tbody> </table> <p>* When using genomic DNA as template DNA, set it to 5 min. † The optimal annealing temperature depends on the melting temperature of the primers.</p>				Step	Temperature	Time	Cycles	Pre-denaturation	95°C	1-5 min*	1 cycle	Denaturation	95°C	30 sec		Annealing	45-65°C†	30 sec	25-35 cycles	Extension	72°C	0.5-1 min/kb		Final extension	72°C
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3	 Analyze with gel electrophoresis	3. Perform the reaction under the following conditions.																									
		4. After the reaction, maintain the reaction mixture at 4-8°C. The samples can be stored at -20°C until use. 5. Load samples on agarose gel with adding a loading-dye mixture (Cat. No. C-9029, not provided) and perform gel electrophoresis for analysis.																									
	 Option	<ul style="list-style-type: none"> If primer's T_m value is more than 65°C or PCR product size is more than 5 kb, follow the conditions as below. <table border="1"> <thead> <tr> <th>Step</th> <th>Temperature</th> <th>Time</th> <th>Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td>95°C</td> <td>1-5 min</td> <td>1 cycle</td> </tr> <tr> <td>Denaturation</td> <td>95°C</td> <td>30 sec</td> <td></td> </tr> <tr> <td>Annealing/Extension</td> <td>68°C</td> <td>1 min/kb</td> <td>25-35 cycles</td> </tr> <tr> <td>Final extension</td> <td>68°C</td> <td>3-5 min</td> <td>1 cycle</td> </tr> </tbody> </table>				Step	Temperature	Time	Cycles	Pre-denaturation	95°C	1-5 min	1 cycle	Denaturation	95°C	30 sec		Annealing/Extension	68°C	1 min/kb	25-35 cycles	Final extension	68°C	3-5 min	1 cycle		
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