

[Cat. No.] Please refer to the **Ordering Information**

Introduction

AccuTarget™ qPCR Screening kit can quantify mRNA gene expression levels by screening the genes with intercalating dyes using real-time PCR. This product contains 92 genes including 84 primer sets and 8 reference genes in lyophilized form in a single plate. This product is divided into three standardized sizes (10X, 20X, 30X) and is convenient to use depending on experimental conditions. The primers were designed using accurate bioinformatics tools in accordance with the MIQE (Minimum Information for Publication of Quantitative Real-Time PCR Experiments) guidelines*. Each well of plate you order come as lyophilized primers (forward and reverse) with a patented stabilizer that maintains full activity for over one month at room temperature, and 2 years in the freezer. Simply use it by dissolving primers and suspending it to your qPCR plate.

* Bustin, S.A., et al. 2009. The MIQE Guidelines: Minimum Information for Publication of Quantitative Real-Time PCR Experiments, Clinical Chemistry 55:4, 611-622.

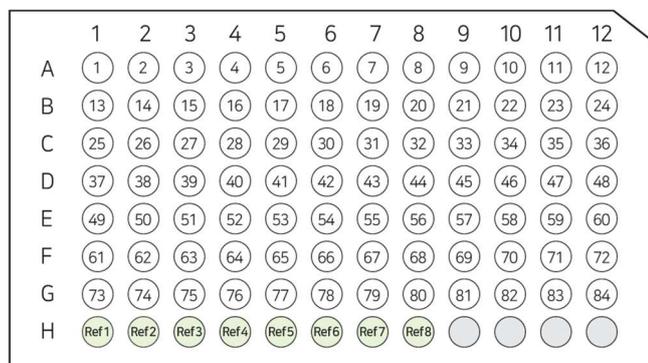


Figure 1. Example of qPCR plate layout.

Features & Benefits

- Professional design: Design the primers by considering all the necessary conditions for qPCR such as target specificity, Tm value, and the size of PCR products.
- Accurate, Cost-saving service: We provide primer sets validated by undergoing target-specific amplification test through meticulously designed algorithms and utilizing BLAST to reduce the time and cost caused by wrong designs.
- Convenience: Easy to use, according to the purpose of the experiment by providing standardized sizes (10X, 20X, 30X) with already pipetted contents for convenience.

Components

Components	Amount
AccuTarget™ qPCR Screening Kit (96 deep-well plate)	1 plate

Storage

- This product is lyophilized and shipped at ambient temperature.
- Store at ambient temperature (15-20°C) without direct sunlight for long term storage. Lyophilized primers are thermo-stable, but once dispensed, primers should be stored at -20°C and repeated freeze and thaw cycles (more than once) are not recommended.

Online Resources



Korean



English

Visit our **product page** for additional information and protocols

Ordering Information

A. Order details

Description		Cat. No.
AccuTarget™ Human qPCR Screening kit	10X	SH-0000-10
	20X	SH-0000-20
	30X	SH-0000-30
AccuTarget™ Mouse qPCR Screening kit	10X	SM-0000-10
	20X	SM-0000-20
	30X	SM-0000-30

* Note: Each of primer is supplied with 10X, 20X, 30X in the well of the plate.

B. Custom order

Description		Cat. No.
AccuTarget™ qPCR Screening Kit- Pathway Custom Kit (Modify the plate design with 1-5 genes)	10X	SPC-0001-10
	20X	SPC-0001-20
	30X	SPC-0001-30
AccuTarget™ qPCR Screening Kit- Whole Custom Kit (Modify the plate design into two more pathway)	10X	SWC-0001-10
	20X	SWC-0001-20
	30X	SWC-0001-30

* Note: Each of primer is supplied with 10X, 20X, 30X in the well of the plate.

Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

Explanation of Symbols



Caution



Consult Instructions For Use

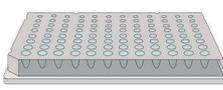


Do not Re-use



Use-by Date

Experimental Procedures

Steps		Procedure Details																																	
1	 Preparation of primers	<p>1. Dissolve primers with nuclease-free water as described in following table.</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 33%;">Concentration*</th> <th style="width: 33%;">Nuclease-free water</th> <th style="width: 33%;">Final concentration</th> </tr> </thead> <tbody> <tr> <td style="text-align: center;">10X</td> <td style="text-align: center;">50 µl</td> <td style="text-align: center;">3 pmol/µl</td> </tr> <tr> <td style="text-align: center;">20X</td> <td style="text-align: center;">100 µl</td> <td style="text-align: center;">3 pmol/µl</td> </tr> <tr> <td style="text-align: center;">30X</td> <td style="text-align: center;">150 µl</td> <td style="text-align: center;">3 pmol/µl</td> </tr> </tbody> </table> <p>* Note: The concentrations (10X, 20X, 30X) are standards set based on the amount used when the reaction is performed with a final volume of 50 µl.</p> <p>2. Spin down the plate and resuspend lyophilized primers with nuclease-free water completely by vortexing.</p>	Concentration*	Nuclease-free water	Final concentration	10X	50 µl	3 pmol/µl	20X	100 µl	3 pmol/µl	30X	150 µl	3 pmol/µl																					
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<p>Example of Real-time PCR Protocol</p> <ul style="list-style-type: none"> When performing real-time PCR, it is compatible with any other products, but we recommend BIONEER's 2X <i>GreenStar</i>™ qPCR Master Mix with following protocols. 																																			
2	 Preparation of reaction mixture	<p>3. Add template DNA, primers, 80X ROX dye (optional), nuclease-free water, and <i>AccuPower</i>® 2X <i>GreenStar</i>™ qPCR Master Mix (Cat. No. K-6251, not provided) into real-time PCR plate (not provided) to make a total volume of 20 µl or 50 µl (recommended).</p> <ul style="list-style-type: none"> Preparation of reaction mixture <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 60%;">Components</th> <th style="width: 20%;">20 µl reaction</th> <th style="width: 20%;">50 µl reaction</th> </tr> </thead> <tbody> <tr> <td><i>AccuPower</i>® 2X <i>GreenStar</i>™ qPCR Master Mix</td> <td style="text-align: center;">10 µl</td> <td style="text-align: center;">25 µl</td> </tr> <tr> <td>Template DNA (5 pg-100 ng)</td> <td style="text-align: center;">Variable</td> <td style="text-align: center;">Variable</td> </tr> <tr> <td>qPCR primer (3 pmol/µl)</td> <td style="text-align: center;">2 µl</td> <td style="text-align: center;">5 µl</td> </tr> <tr> <td>(Optional) 80X ROX dye</td> <td style="text-align: center;">0.25-2 µl</td> <td style="text-align: center;">0.625-5 µl</td> </tr> <tr> <td>Nuclease-free water</td> <td style="text-align: center;">Variable</td> <td style="text-align: center;">Variable</td> </tr> <tr> <td>Total volume</td> <td style="text-align: center;">20 µl</td> <td style="text-align: center;">50 µl</td> </tr> </tbody> </table> <p>* Note: ROX dye is used for normalization of intensity by background subtraction. The use of ROX dye is recommended for Applied Biosystems 7500 Real-Time PCR System (Applied Biosystems), but not required for <i>Exicycler</i>™ 96 Real-Time PCR System (BIONEER), and CFX96 Real-Time PCR System (Bio-Rad).</p> <p>4. Seal real-time PCR plate with adhesive optical sealing film (Cat. No. 3111-4110, not provided).</p> <p>5. Mix the reaction mixture by vortexing, and briefly spin down.</p>	Components	20 µl reaction	50 µl reaction	<i>AccuPower</i> ® 2X <i>GreenStar</i> ™ qPCR Master Mix	10 µl	25 µl	Template DNA (5 pg-100 ng)	Variable	Variable	qPCR primer (3 pmol/µl)	2 µl	5 µl	(Optional) 80X ROX dye	0.25-2 µl	0.625-5 µl	Nuclease-free water	Variable	Variable	Total volume	20 µl	50 µl												
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3	 Real-time PCR	<p>6. Perform the reaction under the following conditions.</p> <p>6-1. PCR condition (2-Step)</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 25%;">Step</th> <th style="width: 25%;">Temperature</th> <th style="width: 25%;">Time</th> <th style="width: 25%;">Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">1-15 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">3-15 sec</td> <td rowspan="2" style="text-align: center;">40-45 cycles</td> </tr> <tr> <td>Annealing & Extension</td> <td style="text-align: center;">58°C</td> <td style="text-align: center;">5-30 sec</td> </tr> </tbody> </table> <p>* Note: Users can adjust the protocol according to their instrument and template DNA sequences to get optimal results.</p> <p>6-2. PCR condition (3-Step)</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <thead> <tr> <th style="width: 25%;">Step</th> <th style="width: 25%;">Temperature</th> <th style="width: 25%;">Time</th> <th style="width: 25%;">Cycles</th> </tr> </thead> <tbody> <tr> <td>Pre-denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">1-15 min</td> <td style="text-align: center;">1 cycle</td> </tr> <tr> <td>Denaturation</td> <td style="text-align: center;">95°C</td> <td style="text-align: center;">3-15 sec</td> <td rowspan="3" style="text-align: center;">40-45 cycles</td> </tr> <tr> <td>Annealing</td> <td style="text-align: center;">58°C</td> <td style="text-align: center;">5-30 sec</td> </tr> <tr> <td>Extension</td> <td style="text-align: center;">72°C</td> <td style="text-align: center;">30 sec</td> </tr> </tbody> </table> <p>7. After the reaction is completed, analyze the results.</p>	Step	Temperature	Time	Cycles	Pre-denaturation	95°C	1-15 min	1 cycle	Denaturation	95°C	3-15 sec	40-45 cycles	Annealing & Extension	58°C	5-30 sec	Step	Temperature	Time	Cycles	Pre-denaturation	95°C	1-15 min	1 cycle	Denaturation	95°C	3-15 sec	40-45 cycles	Annealing	58°C	5-30 sec	Extension	72°C	30 sec
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