

[Cat. No.] **K-7250, K-7260, K-7270**

## Introduction

AccuRapid™ Protein Expression Kit is designed for *in vitro* transcription and translation from target DNA, which produces recombinant proteins in a cell-free system. This is why it is called a cell-free protein expression. This kit is composed of *E. coli* extract and Master mix, and these are used with a template DNA bearing a gene of interest (either plasmid or PCR product). rNTPs and T7 RNA polymerase are used to synthesize mRNA from a template DNA. And ribosomes, tRNAs, amino acids, and etc. are required for a translation step to synthesize recombinant proteins. These materials are supplied by optimized *E. coli* extract and Master mix. AccuRapid™ Protein Expression Kit is based on the T7 expression system.

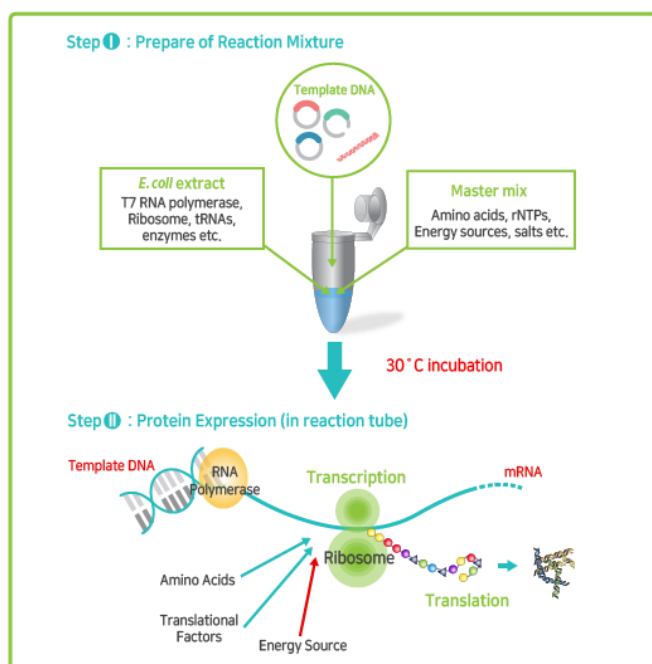


Figure 1. Workflow for protein expression

## Features & Benefits

- Convenient: Includes all necessary components for transcription and translation.
- Rapid: Synthesizes target proteins quickly (within 3 hrs) and economically.
- Flexible: Synthesizes proteins from various types of DNA (plasmid or PCR product).
- Advanced expression technologies: Expression of proteins (cell-toxic proteins, antibodies, membrane proteins, viral proteins, etc.) difficult to be done in the existing *in vivo* technology is made possible.

## Components

K-7250	K-7260	K-7270
AccuRapid™ Cell-Free Protein Expression Kit	AccuRapid™ Midi Protein Expression Kit	AccuRapid™ Maxi Protein Expression Kit

Master mix	0.54 ml	1.2 ml x 2 ea	4.8 ml
<i>E. coli</i> extract	0.3 ml	0.28 ml x 5 ea	2.8 ml
DEPC DW	1 ml	1 ml x 5 ea	3 ml
Positive Control DNA	5 µl	16 µl	10 µl

\* **Note:** For research use only. Not for use in diagnostic or therapeutic procedures.

## Specifications

	K-7250	K-7260	K-7270
Reactions	45 µl x 24 rxns	1 ml x 5 rxns	10 ml x 1 rxn
Expression	Yes (T7 system, Batch type)		
Purification	No		
Target protein size	≤ 150 kDa		
Protein Yield	≤ 300 µg/ml		

## Storage

Store at a temperature between -70°C and -20°C.

## Online Resources



Korean



English

Visit our **product page** for additional information and protocols.

## Ordering Information

Description	Cat. No.
AccuRapid™ Cell-Free Protein Expression Kit	K-7250
AccuRapid™ Midi Protein Expression Kit	K-7260
AccuRapid™ Maxi Protein Expression Kit	K-7270

## Notice

BIONEER corporation reserves the right to make corrections, modifications, improvements and other changes to its products, services, specifications or product descriptions at any time without notice.

## Explanation of Symbols



Batch Code



Catalog Number



Caution



Consult Instructions For Use



Contains Sufficient for <n> tests



Do not Re-use



Manufacturer



Research Use Only




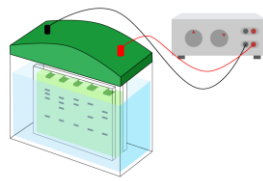


Temperature Limitation



Use-by Date

## Experimental Procedures

Steps		Procedure Details																																							
<b>Preparation of template DNA</b> <ul style="list-style-type: none"> <li>A plasmid or a linear DNA (PCR products) can be used as a template DNA.</li> <li>The template DNA must include a T7 promoter, a ribosomal binding site (RBS), a T7 terminator, and a 6x histidine tag at either N- or C-terminal.</li> </ul>																																									
<b>Cell-free protein expression</b>																																									
1	 <p><b>Thawing materials</b></p>	1. Thaw Master mix, <i>E. coli</i> extract, DEPC DW, and Positive Control DNA on ice. <b>* Note:</b> The pBIVT-AcGFP of about 3.8 kb size is provided as a Positive Control DNA, which has a molecular weight of about 28 kDa.  2. Briefly spin down them and then place the tube on ice. <b>* Note:</b> Make sure that the Master mix and <i>E. coli</i> extract are evenly resuspended before use (be careful not to create bubbles in the extract).																																							
2	 <p><b>Preparation of reaction mixture</b></p>	3. Prepare the reaction mixture (one reaction). <ul style="list-style-type: none"> <li>Preparation of reaction mixture</li> </ul> <table border="1"> <thead> <tr> <th rowspan="2">Components</th> <th rowspan="2">Negative</th> <th rowspan="2">Positive</th> <th>K-7250</th> <th>K-7260</th> <th>K-7270</th> </tr> <tr> <th>45 µl reaction</th> <th>1 ml reaction</th> <th>10 ml reaction</th> </tr> </thead> <tbody> <tr> <td>Master mix</td> <td>21 µl</td> <td>21 µl</td> <td>21 µl</td> <td>0.47 ml</td> <td>4.7 ml</td> </tr> <tr> <td><i>E. coli</i> extract</td> <td>12 µl</td> <td>12 µl</td> <td>12 µl</td> <td>0.27 ml</td> <td>2.7 ml</td> </tr> <tr> <td>Template DNA</td> <td>-</td> <td>1 µl</td> <td>Variable</td> <td>Variable</td> <td>Variable</td> </tr> <tr> <td>DEPC DW</td> <td>12 µl</td> <td>11 µl</td> <td>Variable</td> <td>Variable</td> <td>Variable</td> </tr> <tr> <td>Total volume</td> <td>45 µl</td> <td>45 µl</td> <td>45 µl</td> <td>1 ml</td> <td>10 ml</td> </tr> </tbody> </table> <p><b>* Note:</b> Please refer to our website (<a href="http://www.bioneer.com">www.bioneer.com</a>) for additional information about template DNA amount.</p> 4. Gently mix the reaction mixture by tapping or pipetting.	Components	Negative	Positive	K-7250	K-7260	K-7270	45 µl reaction	1 ml reaction	10 ml reaction	Master mix	21 µl	21 µl	21 µl	0.47 ml	4.7 ml	<i>E. coli</i> extract	12 µl	12 µl	12 µl	0.27 ml	2.7 ml	Template DNA	-	1 µl	Variable	Variable	Variable	DEPC DW	12 µl	11 µl	Variable	Variable	Variable	Total volume	45 µl	45 µl	45 µl	1 ml	10 ml
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3	 <p><b>Incubation for protein expression</b></p>	5. Incubate the reaction mixture at 30°C for 3 hrs in a water bath or a heat block.  6. Briefly spin down the reaction mixture.																																							
<b>Identification of protein expression</b>																																									
4	 <p><b>Analysis with SDS-PAGE</b></p>	7. Analyze the mixture using SDS-PAGE, western blot, or bioactivity assay.  ex) Protocol for SDS-PAGE. <table border="1"> <tbody> <tr> <td>Expression sample</td> <td>5 µl</td> </tr> <tr> <td>4X Loading dye</td> <td>5 µl</td> </tr> <tr> <td>Distilled water (D.W.)</td> <td>10 µl</td> </tr> <tr> <td><b>Total volume</b></td> <td><b>20 µl</b></td> </tr> </tbody> </table> <ul style="list-style-type: none"> <li>- Denaturize at 95°C for 5 min and then load 5 µl of each sample on the SDS-PAGE gel.</li> <li>- Run SDS-PAGE.</li> <li>- Stain the gel with Coomassie Blue R-250.</li> </ul>	Expression sample	5 µl	4X Loading dye	5 µl	Distilled water (D.W.)	10 µl	<b>Total volume</b>	<b>20 µl</b>																															
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