

# AccuPrep® Universal RNA Extraction Kit (K-3140, K-3141)

### 1 Before You Begin

Add 10 μl β-mercaptoethanol per 1 ml RB Buffer.
Prepare additional ethanol(80% and 100%) that is not included.

### Cultured Cell Collection

- 1) Cells grown in suspension:
  - Count the cell number and centrifuge the cultured cells ( $10^4 \sim 10^\circ$ ) at 300 x g for 5 min. Discard supernatant and go to step III. RNA Extraction from Cultured Cell.
- 2) Cells grown in a monolayer. There are 2 different ways to collect cells grown in a monolayer.
  - a. Direct cell lysis from culture dish: Completely remove cell culture medium and go to step III. RNA Extraction from Cultured Cell.
  - b. Harvesting cells with trypsin: Remove cell culture medium and wash the monolayer with DPBS. Add 0.1%~0.25% typsin to the washed cell monolayer. When the cells are detached, add cell culture medium to inactivate typsin. Transfer the cells into a RNase-free tube and centrifuge at 300 x g for 5 min. Discard supernatant carefully and go to step III. RNA Extraction from Cultured Cell.

## INA Extraction from Cultured Cell

- 1) Add 400 µl of RB Buffer to the cell pellet and mix by vortex mixer.
- 2) Add  $300\,\mu l$  of ethanol (80%) and mix immediately by using pipette.
- 3) Transfer the sample to a Binding column in a 2 ml collection tube.
- 4) Close the lid and centrifuge at  $\geq$  14,000 rpm for 20 sec.
- 5) Discard the flow-through from the collection tube and reuse the collection tube.
- 6) Add 700 μl of RWA1 Buffer without wetting the rim, close the tube, and centrifuge at 14,000 rpm for 20 sec.
- 7) Discard the solution from the collection tube and reuse the collection tube.
- 8) Add 500  $\mu l$  of RWA2 Buffer without wetting the rim, close the tube, and centrifuge at 14,000 rpm for 20 sec.
- 9) Discard the solution from the collection tube and reuse the collection tube.
- Add 500 µl of RWA2 Buffer without wetting the rim, close the tube, and centrifuge at 14,000 rpm for 2 min.
- 11) Discard the solution from the collection tube and reuse the collection tube.
- 12) Centrifuge once more at **14,000 rpm** for **1 min** to completely remove ethanol, and check that there is no droplet clinging to the bottom of



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binding column tube.

- 13) Transfer the Binding column tube to a new 1.5 ml tube for elution, add 50~200 μl of ER Buffer onto Binding column tube, and wait for at least 1 min at RT (15~25°C).
- 14) Centrifuge at 10,000 rpm for 1 min to elute.

#### NA Extraction from Plant Tissue

- Grind Sample (up to 100 mg) under liquid nitrogen. Transfer the tissue powder to an appropriately sized tube and add 500 μl of RB Buffer to a maximum of 100 mg tissue powder and vortex vigorously.
- 2) Incubate at 60°C for 1~3 min.
- 3) Centrifuge at full speed for 2 min.
- 4) Transfer the supernatant, aqueous phase to a new microcentrifuge tube.
- 5) Add 0.5 sample volume of ethanol (96~100%) and mix immediately by using pipette.
- 6) Go to **step 3** of "RNA Extraction from Cultured Cell" in page 1 and follow the instructions accordingly.

### **(V)** RNA Extraction from Animal Tissue

- 1) Homogenize the sample (20~30 mg) with a homogenizer, place them in a new 1.5 ml tube, and add **500 μl** of **RB Buffer**.
- 2) Centrifuge the lysate for 3 min at full speed, and transfer the supernatant to a new 1.5 ml tube.
- Add 200 μl of ethanol (96 ~100%) and mix immediately by using pipette.
- 4) Go to **step 3** of "RNA Extraction from Cultured Cell" in page 1 and follow the instructions accordingly.

<sup>%</sup> For more information, please visit www.bioneer.com and refer to the User's Guide of this kit.