MagListo™ 5M PCR/Gel Purification Kit (K-3627)

Ⅰ Before You Begin

1) Prepare the absolute isopropanol and absolute ethanol (not provided in the kit).
2) Gels containing ≤1% agarose for the purification are required.
3) Ensure to add **29 mL** of **absolute isopropanol** to **PB Buffer** before the purification.

Ⅱ Fragment DNA purification from PCR product sample

1) Add **5 volumes** of **PB Buffer** to the PCR product and mix thoroughly by vortexing. (e.g., For 20 µl of PCR product, add 100 µl of PB Buffer in 1.5 ml tube.)
2) Add **100 µl** of **Magnetic Nano Beads Solution** and mix thoroughly by vortexing. **(Note)** Vortex mix the Magnetic Nano Beads Solution before use.
3) Place the tube in ***MagListo*™-2 Magnetic Separation Rack**. The *MagListo*™ stand is attached with the magnet plate. Then invert the tube 3~4 times gently.
4) Without removing the tubes from the rack, carefully pour the supernatant out and remove the remaining supernatant using the paper towel by blotting.
5) Detach the magnet plate from the stand. Add **700 µl** of **W2 Buffer** to the tube and mix thoroughly by vortexing.
6) Attach the magnet plate to the stand and invert the tube 3~4 times gently until the beads are tightly bound to the magnet.
7) Remove the supernatants without removing the tubes from the rack.
8) Detach the magnet plate from the stand. Add **700 µl** of **Absolute ethanol**, and mix thoroughly by vortexing, then repeat the above steps 6~7.
9) Detach the magnet plate from the stand. Completely dry the beads with the tube opened at 60°C using the Heating Block for 10 min. Remove the remaining supernatant using the pipette.
9) Add **30~50 µl** of **EA Buffer** and mix thoroughly by vortexing. Incubate the tube at 60°C for more than 1 min.
10) Attach the magnet plate to the stand and carefully transfer the supernatant containing the nucleic acid to a sterile microcentrifuge tube.

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Ⅲ Fragment DNA purification from Agarose gel

1) Visualize the band in agarose gel stained with any nucleic acid staining chemicals and cut the gel around the DNA band of interest using a scalpel blade.
2) The maximum amount of gel slice in each sample is **approximately 400 mg**.
   Add **3 volumes** of **FB Buffer** to the gel slice (e.g., For 200 mg gel slice, add 600 µl of FB Buffer).
   (Note) Ensure the FB Buffer color remains yellow.
3) Incubate the sample at **60°C** for **10 min** and invert the tube every 2~3 min during the incubation to mix.
   (Note) Ensure complete gel dissolution.
4) Add **100 µl** of **Magnetic Nano Beads Solution** and mix thoroughly by vortexing.
   (Note) Vortex mix the Magnetic Nano Beads Solution before use.
5) Place the tube in **the rack** in which the stand is attached with the magnet plate and invert the tube 3~4 times gently.
6) Without removing the tubes from the rack, carefully pour the supernatant out and remove the remaining supernatant using the paper towel by blotting.
7) Detach the magnet plate from the stand. Add **700 µl** of **FWM1 Buffer** to the tube, and mix thoroughly by vortexing.
8) Attach the magnet plate to the stand and invert the tube 3~4 times gently until the beads are tightly bound to the magnet.
9) Remove the supernatants without removing the tubes from the rack.
10) Detach the magnet plate from the stand. Add **700 µl** of **W2 Buffer** to the tube and mix thoroughly by vortexing. Repeat the above steps 8~9.
11) Detach the magnet plate from the stand. Add **700 µl** of **Absolute ethanol** to the tube, mix thoroughly by vortexing, and repeat the above steps 8~9.
12) Detach the magnet plate from the stand. Completely dry the beads with the tube opened at 60°C using the Heating Block for 10 min. Remove the remaining supernatant using the pipette.
13) Add **30~50 µl** of **EA Buffer** and mix thoroughly by vortexing. Incubate the tube at 60°C for more than 1 min.
14) Attach the magnet plate to the stand and carefully transfer the supernatant containing the nucleic acid to a sterile microcentrifuge tube.

※ For more information, please visit www.bioneer.com and refer to the User's Guide of this kit.

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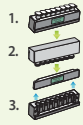
Step	Image	Description
Binding		Add **PB Buffer** to the PCR product and mix. - Add **5 volumes** of **PB buffer** to the PCR product. (ex. For 20 µl of PCR product, add 100 µl of PB buffer)
		Add **100 µl** of **Magnetic Nano Bead** and mix
		Follow these 3 Magnet Attach/Detach Steps 1. Attach the magnet 2. Discard the supernatant 3. Detach the magnet
1st Wash		Add **700 µl** of **W2 Buffer** and mix
		Repeat the above "magnet attach/detach step" (step **1,2 and 3**)
2nd Wash		Add **700 µl** of **Absolute ethanol** and mix
		Repeat the above "magnet attach/detach step" (step **1,2 and 3**)
Drying		Dry the beads with the tube opened at **60°C** for **10 min**
Elution		Add **30~50 µl** of **EA buffer** and mix
		Use the **60°C** Heating block for more than **1 min**

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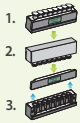
Step	Image	Description
Gel solublization		Add **FB Buffer** to the gel slice - Gel sample: **approx. 400 mg** - Add **3 volumes** of **FB buffer** to the gel slice (e.g., for 200 mg of gel slice, add 600 µl of FB buffer)
		Use the **60°C** Heating block for **10 min** and invert the tube every 2-3 min during incubation
Binding		Add **100 µl** of **Magnetic Nano Bead** and mix
		Follow these 3 Magnet Attach/Detach Steps 1. Attach the magnet 2. Discard the supernatant 3. Detach the magnet
1st Wash		Add **700 µl** of **FWM1 Buffer** and mix
		Repeat the above "magnet attach/detach step" (step **1,2 and 3**)
2nd Wash		Add **700 µl** of **W2 Buffer** and mix
		Repeat the above "magnet attach/detach step" (step **1,2 and 3**)
3rd Wash		Add **700 µl** of **Absolute ethanol** and mix
		Repeat the above "magnet attach/detach step" (step **1,2 and 3**)
Drying		Dry the beads with the tube opened at **60°C** for **10 min**
Elution		Add **30~50 µl** of **EA buffer** and mix
		Use the **60°C** Heating block for more than **1 min**